

The role of oxidized Gly25, Gly29 and Gly33 residues on the interactions of $A\beta_{1-42}$ with lipid membranes

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Abstract

Oxidative stress is known to play an important role in the pathogenesis of Alzheimer's disease. Moreover, it is becoming increasingly evident that the plasma membrane of

neurons plays a role in modulating the aggregation and toxicity of Alzheimer’s amyloid- β peptide, (A β). In this study, the combined and interdependent effects of oxidation and membrane interactions on the 42 residues long A β isoform are investigated using molecular simulations. Hamiltonian replica exchange molecular dynamics simulations are utilized to elucidate the impact of selected oxidized glycine residues of A β 42 on the interactions of the peptide with a model membrane comprised of 70% POPC, 25% cholesterol, and 5% of the ganglioside GM1. The main findings are that, independent of the oxidation state, A β prefers binding to GM1 over POPC, which is further enhanced by the oxidation of Gly29 and Gly33 and reduced the formation of β -sheet. Our results suggest that the differences observed in A β 42 conformations and its interaction with a lipid bilayer upon oxidation originate from the position of the oxidized Gly residue with respect to the hydrophobic sequence of A β 42 involving the Gly29-XXX-Gly33-XXX-Gly37 motif and from specific interactions between the peptide and the terminal sugar groups of GM1.

Keywords

Amyloid- β peptide, Molecular dynamics, Membrane simulations, oxidative stress, GM1, peptide membrane interactions

1 Introduction

Alzheimer’s disease (AD) is a neurodegenerative disorder generally affecting persons 65 years and older (1). With no cure currently available, and the generally increasing life expectancy in most societies, the number of people affected by this disorder is expected to increase in the coming years. Several mechanisms have been proposed as the pathological cause of AD, including genetics, cholinergic, tau and amyloid hypotheses (2–6). Though no hypothesis has been generally accepted, AD brain features substantiate the most widely accepted amyloid

cascade hypothesis (7). In particular, the AD brain is characterized by the presence of senile plaques. The main component of these plaques is the amyloid- β peptide, which is produced by the cleavage of the amyloid precursor protein (APP) by β and γ secretases (8–10). $A\beta$ ranges from 39 to 43 amino acids in length, however, $A\beta_{42}$ is predominant in senile plaques and is known to be more toxic than $A\beta_{40}$ (11–13). The primary structure of $A\beta_{42}$ is shown in Fig. 1. It extends from the unstructured hydrophilic N-terminal region (Asp1 to Lys16) to the hydrophobic C-terminal region (Ala30 to Ala42), and it is linked by central residues (Leu17 to Gly29) that most often forming a turn conformation. The central residues have been shown to play a role in membrane insertion, and sidechain–sidechain interactions through a backbone bend that brings the two β -sheets together (14–18). In addition, the AD brain is commonly characterized by an increase in oxidative stress (19–23). The generation of excess reactive oxygen species (ROS) or the dysfunction of the anti-oxidant system can cause an increase in the amount of ROS present in normal cells and subsequently leads to the oxidative stress observed in AD (24, 25). The ROS are produced either enzymatically (for example to kill invaders in macrophages) or as a side reaction (like respiratory chain), and generally kept at low level but not totally eliminated due to their function. They are necessary to maintain homeostasis in cell and play important role in signaling. The brain seems to be sensitive to oxidative damage upon oxidative stress due to high dioxygen (the final electron acceptor) consumption in the brain; approximately 20% of the total body consumption (26). It is still controversial whether the accumulation of $A\beta$ increases the level of oxidative stress or that the high level of oxidative stress drives $A\beta$ accumulation (6). Studies have shown that $A\beta$ is capable of generating free radicals (27, 28) and shows high affinity to bind metals such as Cu^{2+} , Zn^{2+} (29–31).

Though the physiological role of $A\beta$ is not well understood, it was found that $A\beta$ -membrane interactions are essential for $A\beta$ to fulfill its physiological functions. These functions include protecting the body from infections, repairing leaks in the blood-brain barrier, promoting recovery from injury, and regulating synaptic function (32, 33).

Understanding the effect of membrane composition on the conformation of A β and its interaction with membranes are essential to explain both its physiological function and its role in AD. It was found that the charge of the lipid head group and the structure of the lipid acyl chains determine the transmembrane stability of A β 42 in zwitterionic POPC and DPPC bilayers, and anionic POPG bilayers in MD study by Poojari et al. (17). In another MD study, Poojari et al. found that A β 42 mutation altered both its interactions with neuronal membranes and its aggregation propensity (18). The neuronal membrane plays a key role in modulating A β aggregation (5, 34, 35). It was found that monosialotetrahexosylganglioside (GM1) the most abundant ganglioside in neurons, affects the conformation of A β and its neurotoxicity (36). Moreover, it has been suggested that the initial attachment of amyloid protein to the plasma membrane is first triggered by binding to gangliosides due to their extracellular location (37). Generally, the A β -bound GM1 complex has been identified to be present in the AD brain and it has been found that monomeric A β has a high affinity for GM1 (38, 39). Furthermore, it was found that the GM1-binding domain of A β 42 (A β (1-16)) can inhibit the amyloid pore formation via binding to GM1 and blocking the first step in the pore formation mechanism (a stepwise mechanism controlled by the dual effect of gangliosides and cholesterol) (40). It was also found that A β complex formation is enhanced in lipid bilayers enriched by cholesterol content (41, 42).

The role of A β in oxidative stress related mechanisms of AD progression is still unclear. It was shown that Met35 of A β plays a critical role in this redox process (43, 44). Several studies found that replacing the sulfur atom in Met35 abolished the neurotoxic effect caused by the native peptide (44-47). The neurotoxic fragment A β ₂₅₋₃₅ was observed to lose its neurotoxicity when Met35 was removed (48, 49). It was further hypothesized that the interaction between the proximal residues Gly33 and Met35 in A β 42 accelerates the generation of free radical induced oxidative stress (50, 51). Moreover, it has been proposed that the hydrophobic C-terminal region is the seed for A β aggregation (52). This region includes part of the repeated motif GxxxG spanning residues from Gly25 to Gly37 (53).

Others have shown that Gly33 of this motif plays a key role in A β toxicity (50, 54). It was further reported that modification of Gly33 like mutation not only affects the structure and the hydrophobic surface of the peptide but also affects the neighboring residues and would likely disrupt the interaction between Phe19 and Leu34.

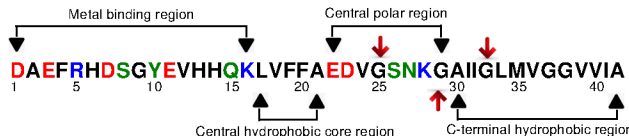


Figure 1: The primary structure of A β 42, showing the acidic residues in red, the basic residues in blue, the hydrophobic residues in black, and the polar residues in green. The residues forming the metal-binding region, the central hydrophobic core, the central polar region, and the C-terminal hydrophobic region are also indicated. Red arrows point to the oxidized glycine residues investigated in this study.

The relationship between glycine residues in the C-terminal region of A β and oxidative stress can be explained by the amyloid radical hypothesis. This hypothesis describes how the C atom of glycine residues is susceptible to the loss of an H-atom upon oxidation of Met35, which in turn causes the formation of a protein backbone radical, stabilized by captodative effect (55, 56), that adopts an extended structure perfect for β -sheet formation (51). In this hypothesis, Gly33 is predicted to be more susceptible to oxidation by methionine-based sulfuranyl free radical due to the close proximity of these residues in the A β 42 primary structure. Several experimental studies have been attempted to explore the effect of radicals on proteins but challenges in locating the center of a radical in protein hinders systematic experimental investigations (57, 58).

Studies of A β peptide often employ shorter A β analogs; however, it is essential to explore the full-length monomeric form of A β for a comprehensive understanding of its toxicity (59, 60). Unfortunately, exploring the full-length of A β is experimentally challenging due to its high propensity to aggregate. Alternatively, MD simulations can provide insight into the conformational dynamics of A β at atomistic resolution, and complements experimental findings (61, 62). Previously, MD simulations have been used to investigate the effect of protein oxidation on protein folding. Owen and coworkers examined the effect of

C_{α} -centered radical formation on the stability of a model helical peptides in different solvent systems (63). Moreover, MD simulations demonstrated the effect of glycine residue radicalization on protein conformation depends both on the protein and the position of the radical (64). The effect of radicalization at Gly25 on A β 42 dimer in solution was further investigated by Liao et. al (65). Although the occurrence of oxidative stress in several neurodegenerative diseases is well-established, there is no computational evidence on the effect of radicalization on peptide-membrane interactions.

Thus, in this study we employ both conventional MD simulations, and Hamiltonian replica exchange molecular dynamic (HREMD) simulations to understand the effect of oxidizing glycine residues (25, 29, and 33) on A β 42 secondary structure and membrane binding. The structure of 1-palmitoyl-2-oleyl-phosphatidylcholine (POPC), monosialotetrahexosyl-ganglioside (GM1) and cholesterol (CHOL) used in this membrane are shown in Fig. 2. The parameters of the glycy radical (GLR) is taken from our previous study (66), and the structure of GLR is shown in Fig. S1 in the Supporting Material. HREMD simulations provide an advantage over MD in that it enhances the conformational sampling of the phase space and overcomes the problem of restricting the system to localized low energy regions of the conformational space (67).

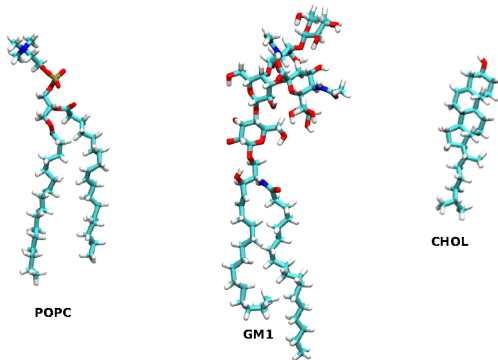


Figure 2: The structure of 1-palmitoyl-2-oleyl-phosphatidylcholine (POPC), ganglioside (GM1), and cholesterol (CHOL) used in this work. The structures are shown in licorice representation colored by name, where oxygen atom is shown in red, nitrogen atom is in blue, hydrogen atom is in white and carbon atom in cyan.

2 Results and Discussion

Understanding the neuropathology of Alzheimer’s disease (AD) is the driving engine for most research in chemical neuroscience and biology. It is thought that the development of Alzheimer’s disease (AD) is related to the interactions between amyloid- β (A β) with neuronal membranes. However, it has also found that the AD brain is characterized by an increase in oxidative stress. This research aims at a better understanding of this complex relationship by examining the effect of oxidizing selected glycine residues in A β 42 on its potential toxicity and interaction with a lipid membrane containing the GM1 ganglioside. That would help toward better understanding of this neurodegenerative disorder and its treatment.

2.1 Effect of bilayer on A β 42

2.1.1 A β 42 Insertion Distance

To see which peptide residues most frequently interacted with the bilayer, we calculated the average distance between the COM of each residues and the average position of each lipid type along the bilayer normal (z-axis). The time-averaged z-position of the phosphorus atom of POPC was used as a reference by setting their average position to zero. The relative position of the hydroxyl oxygen of cholesterol and the COM of the GM1 head group are shown in Fig. 3. On average, the center of mass of the GM1 head groups are above the phosphate group of POPC, while the hydroxy group of cholesterol lay beneath it.

It can be seen in Fig. 3 and Fig. 4 that A β 42-wt and A β 42-GLR29 show similar bilayer interaction profiles, whereas both A β 42-GLR25 and A β 42-GLR33 had similar interaction profile. In the wild-type A β 42 and A β 42-GLR29, Asp1 is closest to the phosphate atoms of the POPC lipids. Its distance from POPC is around 0.5 nm in the former and 0.3 nm in the latter peptide. N-terminal residues Ala2 to Arg5 are the next closest to the bilayer in both cases. The residues in the central polar region (Ser26 to Lys28) were the furthest from the bilayer, whereas residues Ala30, Ile32, Val36, Gly37 from the C-terminal hydrophobic region

of the wt, and residues Lys16, Phe20 of A β 42-GLR29 were the next furthest. Despite these similarities, A β 42-GLR29 stays slightly closer to the bilayer than the wild-type A β 42 does (see Fig. 4). In the case of A β 42-GLR25 and A β 42-GLR33, the former is on average the furthest from the bilayer and this is more pronounced in the N and C-terminal region. In the case of A β 42-GLR25, the central polar region residues Glu22 to Gly29 are the furthest from the bilayer.

To analyze whether the binding of A β 42 to the membrane has an effect on the membrane properties, we first calculated the average bilayer thickness based on the positions of the phosphorus atoms for each simulated system. The average bilayer thickness was found to be about 4.5 nm for all studied systems (see Table S1). This average value was then compared to the local bilayer thickness that was calculated for the snapshots of each trajectory where A β 42 was bound to the membrane. To this aim, we utilized the GridMAT-MD tool (68, 69) to calculate the local membrane thickness averaged over all HREMD snapshots containing at least one atom of A β 42 within 0.5 nm of the lipid bilayer. The resulting 2D plots (Fig. S2) reveal a reduction in the local bilayer thickness to about 4.4 nm or even less upon the interaction of the peptide with the lipid head groups.

2.1.2 A β 42 contacts and hydrogen bond with the bilayer

For a greater understanding of which A β 42 residues associated with the bilayer, the number of contacts between each A β 42 residue and the bilayer for the 4 systems were calculated and normalized in the range of 0 to 1, where 0 means that the residue made no contact with the lipid in question, and 1 stands for residues that always remained in contact with lipid (see Fig. 4).

The N-terminal residues of A β 42-wt made the most contacts with POPC lipids. In this region Asp1 formed the most contacts, followed by Arg5, Glu3, and Ala2 respectively, then His6, Asp7 and Tyr10 from the metal binding region. Fewer contacts, with a normalized contact frequency of less than 0.2, were observed in the central polar and the C-terminal

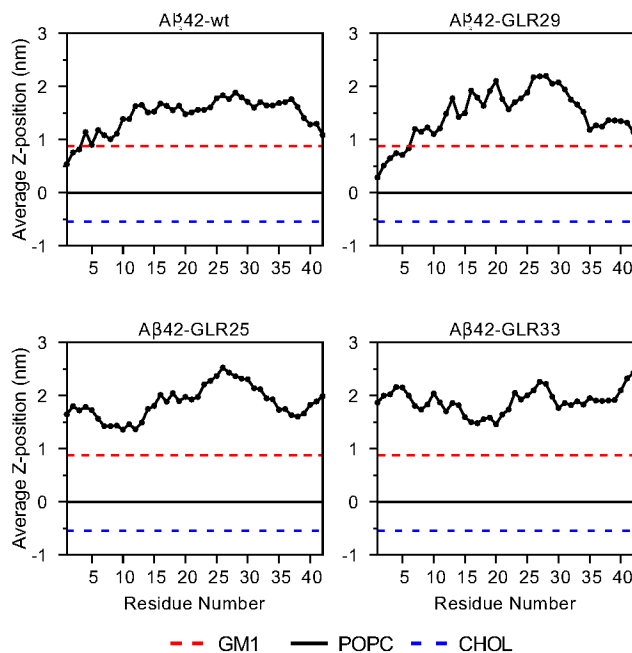


Figure 3: The average insertion distance of each $A\beta 42$ residue with respect to the POPC and GM1 head groups and CHOL hydroxyl group. The head group of GM1, POPC, and CHOL are shown in red, black, and blue respectively.

hydrophobic regions. Similarly, the contacts between $A\beta 42$ -GLR29 residues and POPC were dominated by the N-terminal residues, with a higher membrane affinity shown here in this peptide residues than those in the wild-type. In this region the most contacts were made by Asp1 and the next highest is Ala2, followed by Arg5, Phe4 and Glu3 respectively, and then Tyr10 in the metal binding region. Fewer contacts (a normalized frequency of less than 0.2) were observed in the C-terminal region and almost no contacts were made between the bilayer and the central polar and central hydrophobic regions of the peptide. On the other hand, the contacts between $A\beta 42$ -GLR25 and POPC were dominated by residues from the metal binding region. In this region, contacts with residues Asp1, Arg5, Tyr10, Val12, His13 were most frequent. Fewer contacts, with a normalized frequency of less than 0.5, were observed among the remaining residues in this region and the C-terminal hydrophobic regions. $A\beta 42$ -GLR33 has the lowest affinity for POPC. The most contacts in this peptide were made by residues 13 to 17 from the metal-binding region, with a normalized frequency greater than 0.3.

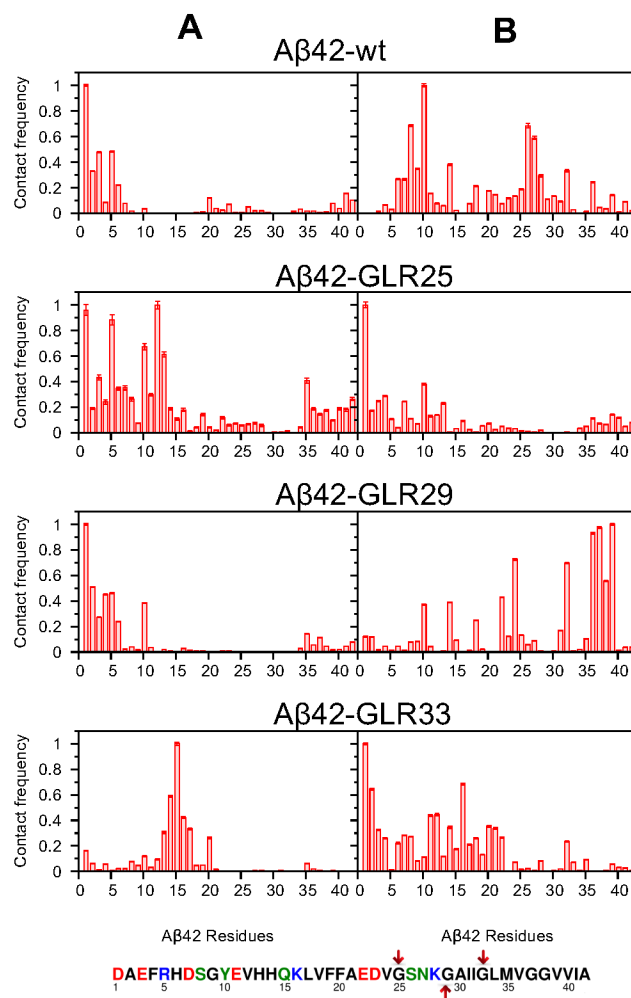


Figure 4: The contact frequency of each $A\beta 42$ residue (and standard deviation of the mean) with POPC (A) and GM1 (B) head groups. The figure shows the normalized contact frequency in the range 0 to 1, such that 1 means the residue always made contacts with the lipid

, and zero means no contacts were made. The sequence of $A\beta 42$ residues is shown below.

Most A β 42 residues showed a propensity to bind GM1 in both the wt and oxidized peptides. However, the contact frequency in the metal-binding region is the highest in A β 42-GLR33, followed by A β 42-wt, A β 42-GLR25, and A β 42-GLR29. The central hydrophobic core and the central polar region had the highest affinity for GM1 in A β 42-GLR33, followed by A β 42-wt, A β 42-GLR29 and A β 42-GLR25 respectively. These contacts are almost abolished in the case of POPC. The C-terminal hydrophobic region showed the highest frequency to bind GM1 in A β 42-GLR29 followed by A β 42-wt, A β 42-GLR33 and A β 42-GLR25 respectively.

Residues Asp1 and Arg5 in the metal-binding region of A β 42-wt and both A β 42-GLR25 and GLR29 formed hydrogen bonds with POPC, as shown in Fig. 5. The propensity to form hydrogen bonds between A β 42 and POPC is higher in the metal-binding region of A β 42-wt and A β 42-GLR29 than in A β 42-GLR25, while it is less than 10% in some residues within the metal-binding region of A β 42-GLR33. On the other hand, no hydrogen bonds formed between POPC and the central polar and C-terminal regions of any of the four A β 42 peptides.

The propensity of hydrogen-bonds to form with GM1 is higher in the case of the oxidized A β 42-GLR25, A β 42-GLR29 and A β 42-GLR33 than in the wt A β 42. The C-terminal hydrophobic residues of A β 42 tended to form hydrogen bonds with GM1, which was not the case with POPC. The highest hydrogen bond propensity of the C-terminal hydrophobic residues was observed in A β 42-GLR29.

2.1.3 A β 42 bilayer interaction energy

Next, we analyzed whether the contacts between A β 42 residues and the lipid bilayer were driven by electrostatic or Lennard-Jones (LJ) interactions. The strongest Coulombic interactions (Fig. S3) between POPC and A β 42-wt was at Asp1 at $-256 \text{ kJ} \cdot \text{mol}^{-1}$, and the next strongest interacting residue was Arg5, however the interaction strength at Arg5 was only 1/5 of that at Asp1. The Coulombic interactions between A β 42-GLR29 and POPC showed

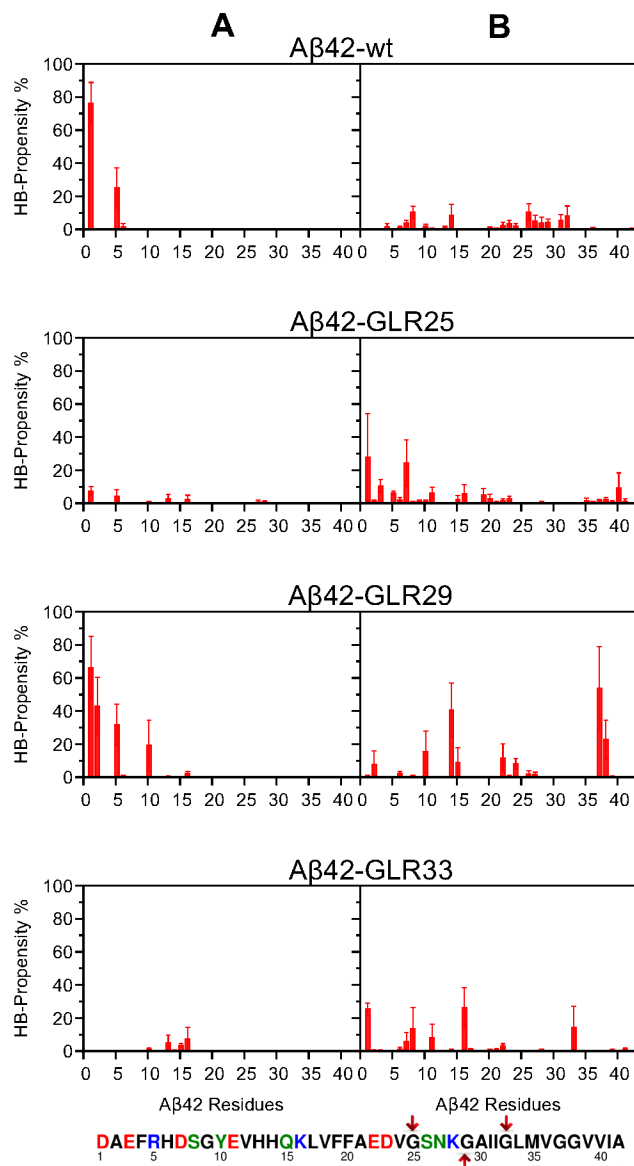


Figure 5: The hydrogen bond propensity (and standard deviation of the mean) of each Aβ42 residue with (A) POPC, and (B) GM1 head groups.

a similar trend as the wt but the interaction energy of Asp1 was weaker, at around -150 $\text{kJ} \cdot \text{mol}^{-1}$, and around -20 $\text{kJ} \cdot \text{mol}^{-1}$ at Tyr10 compared to zero in the wt. The Coulombic interactions between POPC and Aβ42-GLR25 and Aβ42-GLR33 is less than -20 $\text{kJ} \cdot \text{mol}^{-1}$ with residues Asp1 in the former peptide and His13 to Lys16 in the latter peptide. The Coulombic interactions with GM1 was stronger in Aβ42-GLR25 and Aβ42-GLR33 compared to Aβ42-wt and Aβ42-GLR29. The strongest interaction was with Asp1 in Aβ42-GLR25 and with Lys16 in Aβ42-GLR33, and had interaction strengths of -104 , and -60 $\text{kJ} \cdot \text{mol}^{-1}$

respectively. The next strongest interaction between GM1 and A β 42-GLR33 was with Asp1 at $-50 \text{ kJ} \cdot \text{mol}^{-1}$. Overall, the attractive Coulombic interactions with GM1 were lower in magnitude compared to those with POPC, where no single residue had an average interaction energy below $-104 \text{ kJ} \cdot \text{mol}^{-1}$.

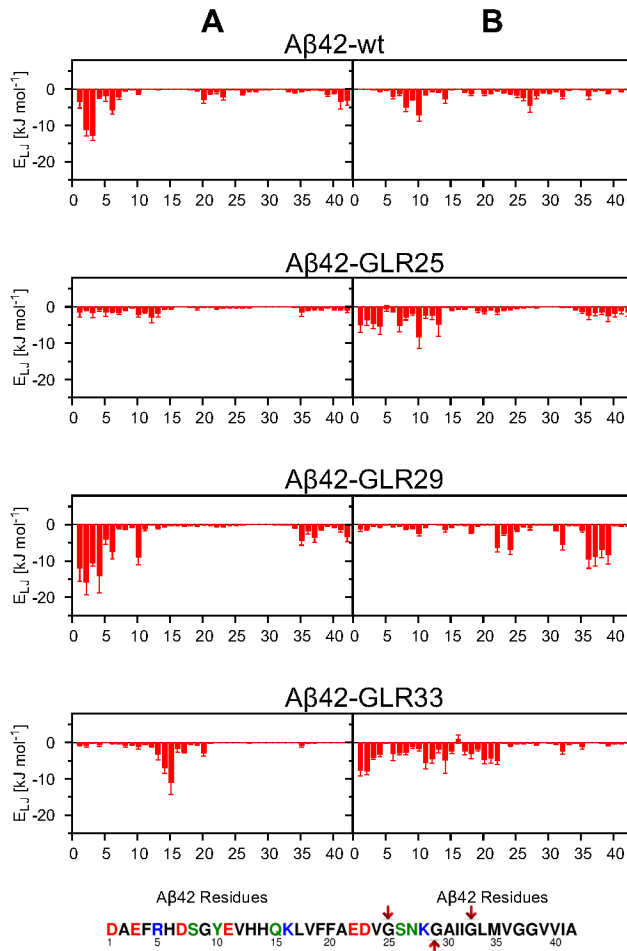


Figure 6: The Lennard-Jones interaction energy of each A β 42 residue (and standard deviation of the mean) between A β 42 and POPC (A) and GM1 (B) head groups.

The profile of the LJ interaction energies (Fig. 6) between each A β 42 peptide and the lipids were very similar to that of their respective contact frequency figure (Fig. 4). For A β 42-wt and A β 42-GLR29 the strongest LJ interactions with POPC were in the N-terminal region of the peptides, and averaged between -10 and $-13 \text{ kJ} \cdot \text{mol}^{-1}$. In A β 42-GLR33 the strongest LJ interactions were with residues His13 to Gln15, whereas residues in the central hydrophobic region averaged between -0.7 and $-11 \text{ kJ} \cdot \text{mol}^{-1}$. Similarly, the strongest LJ

interaction with GM1 was on average $-10 \text{ kJ} \cdot \text{mol}^{-1}$ in the C-terminal region of A β 42-GLR29 and both the metal binding and the central hydrophobic regions of A β 42-GLR33. The LJ interactions of A β 42-GLR25 with GM1 averaged less than $-10 \text{ kJ} \cdot \text{mol}^{-1}$ and was dominated by residues from the metal binding and the C-terminal hydrophobic regions. It averaged less than $-5 \text{ kJ} \cdot \text{mol}^{-1}$ in the the case of the wt A β 42.

Furthermore, we examined the effect of solvation on peptide–lipid interactions, to assure that the stronger the peptide–lipid interaction the lesser the peptide is susceptible to solvation effects. To this aim, the time evolution of the number of water molecules within a 0.5 nm shell from the C-atom of the peptide (70, 71) and the minimum distance of the peptide C $_{\alpha}$ atoms from the lipids were calculated and are jointly shown in Fig. S4. A decreasing peptide distance from the lipid (i.e. stronger peptide—lipid interactions) correlates with a reduction in the peptide hydration. Moreover, the highest population of solvating water molecules was present at approximately 3 nm distance of A β from the membrane surface, i.e., the initial distance and it takes around 50 ns for the peptide to move closer to the lipids.

2.2 A β 42 structural properties

2.2.1 A β 42 secondary structure assignment

In the wild-type A β , residues Phe4 to Glu11 displayed β -turn/bend structure. Residues Val12 to Gln15 displayed helical structure. Residues Lys16 to Ala21 showed a mixture of helical and β -turn/bend structure. Residues Glu22 to Val39 showed β -turn/bend structure with residues Ile32, Leu34, Val36 and Val39 displaying a mixture of β -strand/bridge and helical structure. Residues Met35, Val40, Ile41 showed β -strand/bridge.

Similar to what was seen in the wild-type, all oxidized A β peptides showed a high propensity to form a helical structure in the region Glu11 to Ala21 (Fig. 7), with the lowest propensity observed in the case of A β 42-GLR25. However, an increase in helicity was observed in the case of A β 42-GLR33, such that the helical region extended to include residues Asp7 to Ala21 from the central polar and central hydrophobic regions.

The propensity to form β -strand/bridge was the highest in A β 42-GLR25 and A β 42-wt. Few residues in A β 42-GLR29 and A β 42-GLR33 showed β -strand/bridge structure. This includes residues His6, Tyr10 in the former peptide and residues His13, Leu17, Gly29, Ala30, Val40, Ile41 in the latter peptide. On the other hand, all A β 42 peptides showed a higher probability to form β -turn/bend. Interestingly, residues Asp23 to Asn27 in A β 42-GLR25, residues Asn27 to Ile31 in A β 42-GLR29 and residues Ile31 to Leu34 in A β 42-GLR33 did not show any turn/bend structure.

Moreover, the development of the secondary structure content for all simulations as a function of the simulation time was analyzed and shown in Fig. S5. To this aim we utilized the moving window statistics (more about the method can be found in the figure caption) to test the convergence of the secondary structure during the simulation (71, 72). The figure shows good agreement in the overall trend of the β -sheet propensity over different time windows, and the secondary structure propensity of the peptides apart from A β 42-GLR25 converges after 80 ns as no further changes in the β -sheet propensity are observed by the end of the simulations. A β 42-GLR25 is more flexible than the other three peptides, and thus needs more time for the secondary structure to converge. This is in line with the previous findings by Liao et. al. who found that the flexibility of A β 42 increases upon oxidation of Gly25 (65). Nonetheless, within the last 40 ns of the HREMD simulation also this simulation converged as supported by the nearly stable β -sheet propensity for each of the residues.

2.2.2 A β 42 intramolecular hydrogen bond and contact

Hydrogen bonds present within the peptide backbone, the peptide side chains, and between the peptide backbone and side chains that were present for more than 40% of the time were recorded and listed in Table S2. In the wild-type A β 42, backbone-backbone hydrogen bonds were present in the C-terminal region between Ile41 with Gly33 and Leu34, Ile31 with Leu34 and Met35, Gly33 and Met35, Val36 and Val39, which stabilizes the β -sheet structure. They are also present between residues from the metal binding region such as Glu11 with

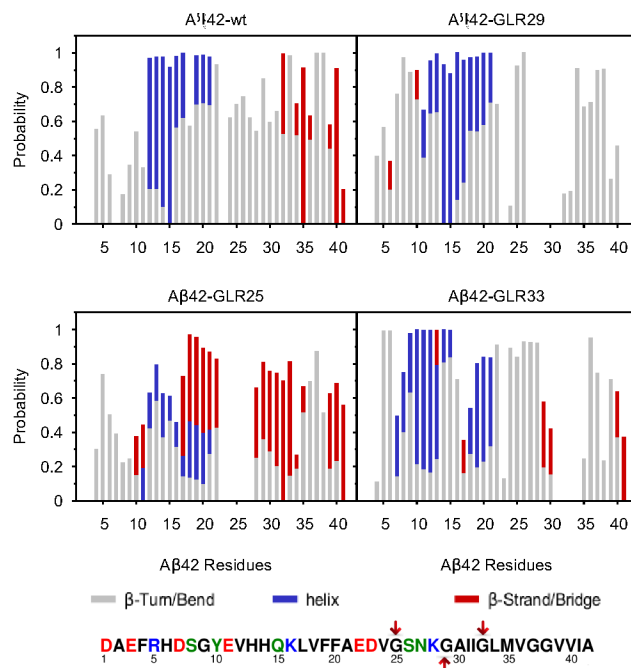


Figure 7: The secondary structure assignment of each A β 42 residue in the case of A β 42-wt, A β 42-GLR25, A β 42-GLR29, and A β 42-GLR33 in a bilayer comprised of 70% POPC, 25% CHOL, 5% GM1. The β -turn/bend is shown in silver, the helix in blue and the β -strand/bridge is shown in red. The figure shows the additive probability of all secondary structure such that the maximum is 1, those residues showing probability lower than 1 form random coil.

His14 and Gln15, Val12 and Lys16, His14 with Leu17 and Val18, Val18 with Ala21 from the central hydrophobic region. In the N-terminal region, backbone-side chain hydrogen bonds formed between Glu3 with Phe4 and Arg5. Moreover, hydrogen bonds formed between the side chain of Glu3 (hydrogen bond acceptor) and the side chain of Arg5 (hydrogen bond donor) from this region. Similarly, a lower number of backbone-backbone hydrogen bonds were present in the A β 42-GLR33 metal binding, and C-terminal regions. On the other hand, A β 42-GLR29, and A β 42-GLR25 contained additional backbone-backbone hydrogen bonds that formed within the central hydrophobic core, within the central polar region, and between the C-terminal and the central polar region in the former, and within the polar region, and between the C-terminal and the central hydrophobic cores of the latter. These bonds exist between residues that either form helices or β -sheets, and such structures are discussed in the next section about the most prevalent A β 42 structures. Residues in the N-terminal region

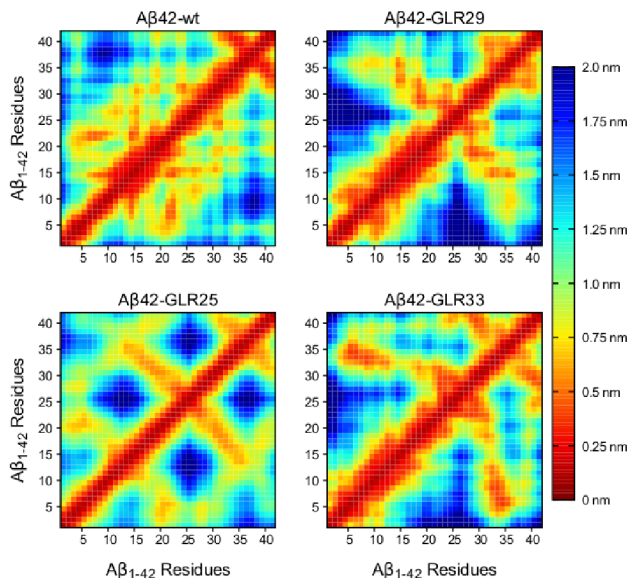


Figure 8: Contact map of each A β 42 residue in the case of A β 42-wt, A β 42-GLR25, A β 42-GLR29, and A β 42-GLR33 in a bilayer compromised of 70% POPC, 25% CHOL, 5% GM1. The color box to the right shows the corresponding distance (in nm) to color present in the contact map.

did not form backbone to backbone hydrogen bonds in any of these peptides.

The backbone-side chain hydrogen bonds formed most often in A β 42-GLR33. However, Phe4 formed hydrogen bond with Glu3 and Arg5 in all peptides except A β 42-GLR25, which did not form any backbone-side chain hydrogen bonds. The hydrogen bonds involving side chains in A β 42-GLR29 were similar to those observed in A β 42-wt as is shown in Table S2. However, A β 42-GLR33 formed an additional hydrogen bond between Ser26 and Asp23, and A β 42-GLR25 did not form side chain to side chain hydrogen bonds.

The contact map between the A β 42 residues is shown in Fig. 8. Regions marked in red indicates residues in close contact (within 0.25 nm of each other) and regions marked in blue stand for residues that display little to no contact. Based on this assignment one can explain the red main diagonal seen in all contact maps, as it represents the contact of a residue with itself and its neighboring residues. Of interest are those islands of red color that spread in almost all contact maps. Considering the contact map of A β 42-GLR25, these light-orange-colored islands are arranged in cross diagonals between residues 28-34 with residues 14-24,

illustrating the formation of β -sheets. Similarly, these diagonals are observed in the other peptides but with some distortion and to a lesser degree in the following order A β 42-GL29 > A β 42-GL33 > A β 42-wt.

2.2.3 The most prevalent A β 42 structures

To substantiate the secondary structure and contact map analysis, we clustered the A β structures using the algorithm developed by Daura et al.. The top cluster of each system is presented in Fig. 9, while the structure in the second and third-largest cluster are shown in Fig. S7-S8. The three highest populated A β 42-wt clusters represented 46.7%, 23.7%, and 9.5% of all structures respectively (Fig. 9A, Fig. S6A, and Fig. S7A). From the N-terminus to the C-terminus, the secondary structure of the largest and second-largest cluster alternated from random coil to turn, followed by helix at Val12 to Leu17, then a turn to anti-parallel β -sheet (on either sides of a turn centered at Gly38) at the C-terminal. There were no β -sheet in the third-largest cluster, however, A β 42 contained coil, turn, and helix. The N-terminus was embedded more deeply into the bilayer in the largest and third-largest clusters, and was least embedded in the second-largest cluster. The three most populated structures found in A β 42-GLR25 represented 33.8%, 18.4%, and 9.4% of all structures respectively are shown in Fig. 9B, Fig. S6B, and Fig. S7B. The β -sheet structure was observed only in the largest and second-largest clusters, however, a much higher β -sheet content can be seen in these clusters. In addition to the β -sheet at C-terminal, a β -sheet was observed at the central hydrophobic and polar region (residues Leu17 to Asp23, Lys28 to Leu34) in the largest cluster, and at the metal binding region (Gly9 to Val12) in the second-largest cluster. A helical structure was only observed in the second and the third largest clusters. The N terminus is closer to the bilayer in the largest cluster, while the the C-terminus is close to the bilayer in both the second and third-largest clusters. In the case of A β 42-GLR29, the three representative structures representing 56.5%, 19.3%, and 3.8% of the total number of structures, respectively, are shown in Fig. 9C, Fig. S6C, and Fig. S7C. No β -sheets observed

in any of the three clusters. However, helical structure was seen in all clusters. Moreover, much more helical content was seen as the helical structure extends to include Glu11 to Glu22, and His13 to Ala21 in the second and the third-largest clusters respectively. The N-terminus is embedded in the bilayer in all three clusters, while the C-terminus is only embedded in the second-largest cluster. Fig. 9D, Fig. S6D, and Fig. S7D shows the three most populated structures in the presence of A β 42-GLR33, which represent 65.6%, 16%, and 5.2% of the total structures respectively. In this case C-terminal β -sheet was observed only in the largest cluster. A helical structure observed in both the largest and second-largest cluster. There were no helices, no β -sheet in the third largest cluster. The N-terminus is close to the bilayer in the second-largest cluster, while both the N and C-terminus residues interact with the solvent in both the largest and the third-largest clusters.

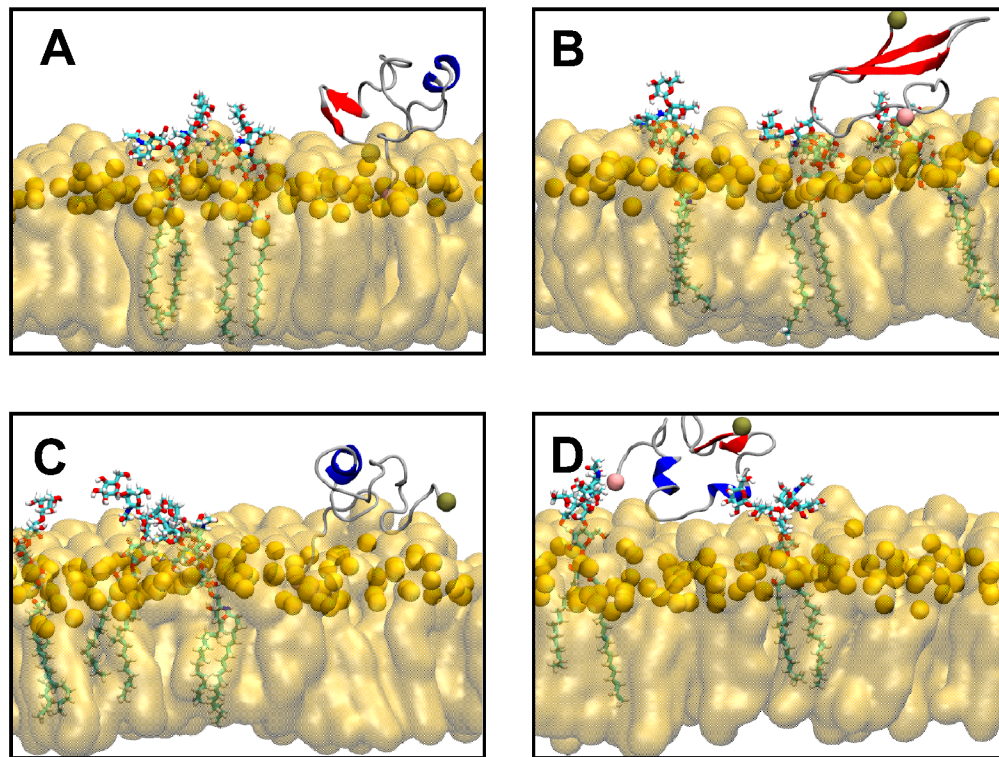


Figure 9: The central structure of the the largest cluster. (A) A β 42-wt (46.7%), (B) A β 42-GLR25 (33.8%), (C) A β 42-GLR29 (56.5%), (D) A β 42-GLR33 (65.6%). In each rendered image the lipids are colored by orange. The phosphate atom of POPC is in orange, the N and C terminals of A β 42 are shown in pink and tan spheres respectively. The protein β sheet is in red, the helix is in blue, coil and turn are shown in silver.

2.3 Discussion

2.3.1 Effect of bilayer on A β 42

The insertion data shows the time-averaged distance between each A β 42 residue and the bilayer. The insertion depth of A β 42-wt > A β 42-GLR29 > A β 42-GLR25 > A β 42-GLR33, as characterized by the average insertion distance (Fig. 3). The detailed interactions of A β 42 residues with the bilayer are shown more quantitatively in the peptide-bilayer contacts Fig. 4. Interactions between A β 42 and the bilayer occur primarily between the N-terminus and the POPC head groups, while the remaining residues make more contact with GM1. A β 42-GLR25 makes the least contact with GM1 as the normalized frequency is generally less than 0.5, which can indicate why it has the tendency to adopt the β -sheet conformation while on membrane surface. As shown in Fig. 4, the contact between peptides and GM1 are ranked in the order of A β 42-GLR29 > A β 42-GLR33 > A β 42-wt > A β 42-GLR25. Contacts observed at the N-terminus especially with Asp1, and Arg5 were driven by hydrogen bonds that formed between the amino acid residues and the lipids. The propensity of hydrogen bonds to form between A β 42 and GM1 are in the order of A β 42-GLR29 > A β 42-GLR33 > A β 42-GLR25 > A β 42-wt. The hydrogen bond propensity with POPC was limited to residues from the N-terminal metal-binding region, with Asp1 and Arg5 as the main hydrogen bond forming residues. This is expected to be the cause that drives the N-terminus to be close to the bilayer head group as can be seen from the insertion data in Fig. 3. Here, A β 42-wt and A β 42-GLR29 showed a higher hydrogen bond propensity with POPC than A β 42-GLR25 and A β 42-GLR33. It is not surprising that A β 42 residues that form a hydrogen bond with the bilayer were also shown to be the strong energetic contribution to interactions between each peptide and the lipid bilayer. The interactions between each A β 42 residue and the lipid bilayer were divided into their Coulombic and Lennard-Jones contributions. Overall, it can be seen from Figs. 6 and S3 that the A β -bilayer interaction are mostly hydrophobic in nature, since most residues interacted with the bilayer via Lennard-Jones interaction.

However, Coulombic interactions played a major role in the case of Asp1 and Arg5 residues in both A β 42-wt and A β 42-GLR29, which is expected due to their tendency to form hydrogen bonds with POPC.

The observation that A β 42 formed most contacts with GM1 can be attributed to the large head group of GM1 containing five sugar groups (Fig. S8) that tend to lay on the membrane surface, thus providing a platform for hydrogen bonding with A β 42 through its sugar head groups (Fig. 4). It should be noted that with 5% GM1 in the bilayer there were only 7 GM1 lipids in each leaflet. Despite this low GM1 content, as can be seen from the most populated clusters (Figs. 9, S6 and S7), A β 42 has a preference to bind to GM1 and in some cases even to two GM1 molecules at the same time due to the possibility of hydrogen bonding to the penta-saccharide GM1 head group. In order to further elucidate the A β 42-GM1 binding, we dissected these interactions into their contributions per sugar ring. The representative snapshot in Fig. S9 and the contact probability in Fig. S10 show that independent of the oxidation state of A β 42, the peptide made most contacts with the sugar groups furthest away from the membrane surface, that is the terminal β -D-Galactose (Gal'), the N-acetyl- β -D-galactosamine (GalNAc), and the N-acetyl- α -neuraminidate (Neu). Apart from A β 42-GLR25, the other three A β 42 variants preferred binding to Neu, while A β 42-GLR25 formed more contacts with Gal'. The contact probability of A β 42-wt and A β 42-GLR25 with GalNAc is also considerably high, leading to some contacts with the adjacent Gal, while A β 42-GLR29 and A β 42-GLR33 formed only few contacts with GalNAc. Fig. S11 confirms that in all cases the contacts between A β 42 and the sugar groups are driven by hydrogen-bond formation, which, however, does not only involve Coulomb but also Lennard-Jones interactions (Fig. S12). The formation of hydrogen bonds of A β 42 with GM1 prevents the peptide from forming intrapeptide hydrogen bonds as needed for secondary structure formation, e.g., the formation of β -sheets and α -helices. This is indeed the case as the numerous snapshots in Figs. 9, S6 and S7 reveal: at the A β 42-GM1 binding sites, no β -sheet or α -helix is found. However, the binding of A β 42 to GM1 keeps A β 42, or at least

parts of it somewhat above the membrane surface (Fig. 3), which in turn encourages β -sheet formation as seen for A β 42-GLR25 in Fig. 9B or helix formation as observed for A β 42-GLR29 and A β 42-GLR33 in Fig. 9C and D. The higher amount of structure formation of membrane-associated A β 42 compared to the solution state can thus be considered to be a consequence of the reduction in conformational flexibility on the membrane surface, while leaving A β 42 enough conformational freedom and also possibilities for intrapeptide hydrogen-bond formation needed for β -sheet or helix formation.

2.3.2 Effect on A β 42 secondary structure

The secondary structure of A β 42 has similar secondary structure elements in each system, particularly in the helical central polar region (Val12 to Ala21). This region remained helical in all peptides, though it is extended to include residues Asp7 to Ala21 in A β 42-GLR33. As evidenced by the membrane contacts, hydrogen bonding, and interaction energies the conformation of A β 42 in the N-terminal region was not affected by the interaction with POPC. This is because A β 42-GLR25 showed higher contact frequency with POPC in the N-terminal compared to A β 42-GLR33, however both peptides showed no tendency to form β -sheets in the N-terminal region. The same is applied to A β 42-GLR29 and A β 42-wt. On the other hand, the interaction with GM1 affected the formation of β -sheets. It was observed that A β 42-GLR29 and A β 42-GLR33 showed the highest number of contacts with GM1 when compared to A β 42-wt and A β 42-GLR25, but these peptides had a lower tendency to form β -sheets, which is in agreement with the finding of Mandel et al. on A β 40 (73). This is also confirmed by the insertion data (Fig. 3) which shows that A β 42-wt and A β 42-GLR25 behave differently with respect to the bilayer, with both peptides having a lower tendency to bind to GM1 (Fig. 4) than A β 42-GLR29 and A β 42-GLR33 does.

The assignment of secondary structure is shown in Fig. 7 and it mirrors what is seen in the most populated clusters Fig. 9. The intra-peptide backbone hydrogen bonds indicate a structure that is very similar to that of a β -sheet in the most populated clusters for these

two systems.

2.3.3 Comparison with other studies

It is known from experimental findings and MD simulations that A β 42 is an intrinsically disordered protein, containing all possible secondary structures (9, 74–77). It has also been shown that the C-terminal β -sheet is the seed for further A β aggregation into β -sheet rich structure (74). Moreover, the importance of the total β -strand content for controlling the aggregation rates was pointed out in an MD study by Man et al. (78). In our study we observed a C-terminal β -sheet in the case of A β 42-wt and A β 42-GLR25 especially in the two regions, Val39-Ile41 and Ala30-Val36. We also found that there is a β -sheet forming propensity in the region of Leu17 to Glu22 and Tyr10 to Glu11 in A β 42-GLR25. However the β -sheet forming propensity in the C-terminal hydrophobic region was very low in the cases of A β 42-GLR29 and A β 42-GLR33, which might be explained by the findings of Fonte et al. (53) and Harmeier et al. (54). Fonte et al. found experimentally that changes in the glycine zipper Gly29-XXX-Gly33-XXX-Gly37 motif prevent the formation of toxic oligomers as they observed a reduction in A β toxicity upon the substitution of Gly37 with leucine (53). It was also found by Harmeier et al. that the substitution of Gly33, which is both at the end of one and at the start of the next GXXXG interaction motif, causes the formation of less toxic oligomers. Based on that, we suggest that the stability of the C-terminal β -sheet in A β 42-GLR25 might be attributed to the fact that Gly25 does not disrupt the zipper motif.

According to our findings, the effect of radicalization of Gly residue on the structure of A β 42 depends on the position of the oxidized Gly residue within the C-terminal hydrophobic region of A β . This is supported by the MD study of Owen and coworkers on three fast folding miniproteins, where it was also found that denaturing effect of the Gly radicals depended on the position of the radical (64). Structural changes in the protein upon Gly oxidation were more pronounced in the α -helix rich protein than the β -sheet rich one due to the flat geometry of the radical (Fig. S1) (64). This may even cause the formation of a β -sheet as

Owen et al. showed in an MD-study of a short helical peptide that underwent a transition to a β -sheet conformation upon radical formation (63).

It has been shown experimentally that $A\beta$ interacts strongly with lipid bilayers composed of phosphatidylcholine head groups (79, 80). In our study we found that this interaction is mediated by hydrogen bonds with the positively charged N-terminal Asp1 and the side chain of Arg5. Furthermore, we found that such tight interactions have no effect on $A\beta$ secondary structure, which is in agreement with the experimental findings that binding of $A\beta$ to PC-containing bilayers does not affect the secondary structure of the $A\beta$ peptide at low concentrations (81, 82). Our results showed how the bulky GM1 head groups covers a large area of the membrane even with only 7 GM1 lipids in each bilayer leaflet, and the peptide is more likely to bind to the GM1 head groups, especially to the terminal saccharide residues, than to the POPC head groups. This agrees with the findings by Manna et al. who concluded that the GM1 head groups act as scaffold for $A\beta$ binding through sugar-specific interactions (83). They further observed the formation of a C-terminal β -hairpin upon binding of $A\beta$ 42 to GM1, which is also in line with our results. However, it should be noted that another study found that binding of $A\beta$ 40 to GM1 head groups induces the formation of a helix on the C-terminal side of the peptide (84). We also observed helix formation upon GM1 binding, which, however, occurred preferentially between residues 10 and 21 of $A\beta$ 42. This includes the region from 17 to 21 that was already identified as helical for $A\beta$ 40 in solution (85). From an experimental study it was followed that low concentrations of GM1, i.e. physiological concentration of GM1 that range from 2–4% of the total lipid content of the membrane (86), enhance the formation of β -sheets, but prevents $A\beta$ oligomerization (87). Other experiments further showed that the higher the concentrations of GM1 (but less physiologically relevant), the higher the β -sheet content of $A\beta$ becomes (88).

3 Methods

3.1 Model System and System Equilibration

The starting structures for the wild-type (wt) and oxidized (Gly25, Gly29, Gly33) A β peptides were obtained from 1 μ s simulations, using the final snapshots of these simulations (Fig. S13). These simulations had begun with the structure of A β 42 as determined by NMR spectroscopy (PDB ID: 1Z0Q) (89). The N- and C-terminals were the free amino (NH 3^+) and carboxyl (COO $^-$) groups, respectively, and thus each peptide carried an overall charge of -3 . The oxidized A β 42 peptides will be henceforth referred to as A β 42-GLR25, A β 42-GLR29 and A β 42-GLR33. The four peptides were placed 3.5 nm above a symmetric membrane composed of 202 POPC, 72 CHOL and 14 GM1 lipids. Each system was solvated with TIP3P water model (90) and neutralized with 17 Na $^+$ ions. In addition, 150 mM of NaCl were added to mimic the physiological concentration of these ions. The exact numbers of each molecules present in each system are listed in Table 1. All interactions among system constituents were described using OPLS-AA force field parameters (66, 91–93). All MD simulations carried out using GROMACS 4.6 simulation package (94).

The systems were energy minimized using the steepest descent algorithm to remove all atomic clashes (95, 96). This was followed by an equilibration under NVT conditions for 1 ns, where the reference temperature of 310 K was regulated with the velocity-rescale thermostat (97) and the time constant set to 0.1 ps. During minimization and initial equilibration stages, the heavy atoms of protein and lipids were subjected to position restraints with a force constant of 1000 kJ/mol $\cdot$ nm 2 . Next, the system was equilibrated under NPT conditions for 10 ns to obtain a pressure of 1.0 bar. The pressure was regulated using semi-isotropic Parrinello-Rahman pressure coupling scheme (98–100) with a time constant of 5 ps and isothermal compressibility of 4.5e-5 bar $^{-1}$. The temperature of 310 K was maintained with Nosé-Hoover thermostat (101–104) with a time constant of 0.5 ps, while the position restraints on the protein and lipids were still on. The particle Mesh Ewald (PME) (105, 106)

method was used to account for the electrostatic interactions within the system. Both the short-range interactions, with a van der Waals cutoff of 1.0 nm (real space), and the long-range (Fourier) electrostatics were used under the periodicity assumption, and the periodic boundary conditions were set in all directions. All bonds were constrained using LINCS algorithm (107).

3.2 Hamiltonian Replica Exchange MD (HREMD) Simulations

For the final production run, the input parameters were the same as those under NPT conditions, except that the position restraints were switched off. To accelerate conformational sampling, we employed the HREMD protocol for the systems described in the previous section using the protocol introduced in (108). System coordinates and input parameters (without restraints) from the NPT equilibration served as the initial input to generate the post-processed topology files required for the HREMD simulations. The system was split into hot ($A\beta$ peptide) and cold (the rest of the system) regions, where the interactions within the hot region and between the hot region and the cold region was enhanced by scaling the force field terms for the proper dihedrals, Lennard-Jones parameters and electrostatic interactions. HREMD simulations were performed using 14 replicas with λ values ranging from 1 to 0.5 and replica exchanges were attempted every 2 ps. Each system was simulated for 200 ns per λ value with the exchange acceptance ratio of 0.13. All simulations were carried out with GROMACS 4.6 patched to PLUMED 2.2. (109).

3.3 Analysis methods

3.3.1 Bilayer-peptide interactions

The analysis of each system began when $A\beta$ was within 0.5 nm of the bilayer. All analysis programs mentioned in this section are included in the GROMACS 2018.2 program package (110–113). The ‘gmj traj’ program was used to measure the insertion distance of $A\beta$ 42

by computing the center of mass (COM) of each residue and the average vertical position of the phosphorus atoms of phospholipids was taken along the z-axis. The ‘gmX mindist’ program was employed to determine the number of contacts between each A β 42 residue and each POPC or GM1 lipid. A contact was recorded when the distance between any two non-hydrogen atoms from the residue and lipid in question was within 0.5 nm. Then the number of contacts were normalized in the range of 0 to 1. The hydrogen bond propensity was determined by the number of times a hydrogen bond was formed between hydrogen bond donating and accepting atoms in A β 42 and each lipid type using the ‘gmX hbond’ program. A hydrogen bond was recorded when the angle between the donor and acceptor bonded hydrogen was between 150 and 180 degrees and the distance between the two atoms was within 0.35 nm. The ‘gmX energy’ program was used to calculate the interactions energy between each A β residue and the head group of POPC or GM1.

Table 1: The molecular composition of the four membrane systems; A β 42-wt, A β 42-GLR25, A β 42-GLR29, and A β 42-GLR33.

Bilayer system	Number of molecules						
	Peptide	Lipids			Ions		Water
	A β 42	POPC	CHOL	GM1	Na ⁺	Cl ⁻	
A β 42-wt	1	202	72	14	69	52	19301
A β 42-GLR25	1	202	72	14	69	52	19289
A β 42-GLR29	1	202	72	14	69	52	19293
A β 42-GLR33	1	202	72	14	69	52	19259

3.3.2 A β 42 structure

The secondary structure of each A β 42 residue was determined using the *define secondary structure program* (do_dssp) (114). To facilitate a clear representation, the data of similar secondary structures are grouped together; β -strand and β -bridge are combined as β -strand/bridge, β -turn and bend as β -turn/bend and helix includes α , π , and 3_{10} helices. Hydrogen bonds within the peptide backbone, between the backbone and side chains, and between the side chains were counted by applying the same method used for counting the

peptide-lipid hydrogen bonds. Representative A β 42 structures were obtained by the ‘gmx cluster’ program using the method of Daura et al. (115) and a cutoff of 0.25 nm for clustering. The conformation and membrane interactions of the central structure of the three largest clusters were rendered using the VMD program (116).

4 Conclusion

Understanding the interplay between oxidative stress and A β neurotoxicity requires exploring the conformation of oxidized A β peptides. Based on our MD simulations of A β 42 and its oxidized variants in interplay with a model membrane bilayer we found that A β 42-GLR25 is potentially as toxic as A β 42-wt, assuming that β -sheet formation in A β is connected to its toxicity (12), whereas A β 42-GLR29 and A β 42-GLR33 showed less β -sheet forming propensity. We also revealed that the sugar moiety of GM1 affects the interaction between A β 42 and the membrane. A β has a high tendency to interact with GM1 (especially A β 42-GLR29 and A β 42-GLR33), and once this happens the propensity of the peptide to form β -sheet is greatly reduced as A β , instead forming intrapeptide hydrogen bonds, interacts with GM1 through hydrogen bonds. Moreover, the interaction with GM1 also reduces the number of contacts and hydrogen bonds that the peptide makes with POPC. On the other hand, the insertion of the peptide into the bilayer is enhanced by its interaction with POPC; A β 42-wt and A β 42-GLR29 showed the highest number of contacts with POPC and therefore these peptides interact most closely with the bilayer. Our results suggest that the differences observed in A β conformation and interaction with the bilayer upon the oxidation of different glycine residues might be attributed in part to the position of these residues within the C-terminal hydrophobic region of A β and its subsequent interaction with GM1. Further studies should test this observation and further determine the role of oxidation in A β -mediated AD toxicity.

5 Author Contributions

All authors made a significant contribution to this manuscript. The contribution of each author is as follows:

H. Fatafta: Analyzed simulation data, prepared figures and presented data, wrote manuscript.

C. Poojari : Designed the study, set-up simulations, wrote manuscript.

A. Sayyed-Ahmad : Provided study funding, wrote manuscript.

B. Strodel: Provided study funding, interpreted simulation data, wrote manuscript.

M. Owen : Designed study, provided study funding, carried out simulations, wrote manuscript.

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Supporting Information Available

The supporting information is available at It includes explanation figures and table that further explain the structure used in this study and enhance our analysis.

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Graphical TOC Entry

